

## 2010 Annual report

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Draths Corporation

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Award Title: Green Synthesis of Phloroglucinol: Exploiting *Pseudomonas fluorescens* and Scale-Up

### Technical Report

#### a. Scientific and Technical Objectives

A new generation of phloroglucinol synthesizing microbe has been evaluated using resin-based extractive fermentation at 1 L scale. Strategies to understand *in vivo* phloroglucinol synthase expression in this genetically engineered *E. coli* were examined. Improving phloroglucinol synthase activity is essential to increase the microbial phloroglucinol synthesis titer and yield. Strategies to identify novel phloroglucinol synthases were therefore explored. It is also believed that expression of phloroglucinol synthase *phlD* gene inside its native *Pseudomonas fluorescens* Pf-5 will likely deliver higher expression level and activity of this gene product. Constructing a phloroglucinol synthesizing *P. fluorescens* was therefore pursued.

#### b. Approach

Coupling metabolic engineering and reaction engineering, improved phloroglucinol synthesizing *E. coli* PG1/pKIT10.080 was evaluated under resin-based extractive fermentor-controlled conditions. Heterologous expression of *P. fluorescens* Pf-5 *phlD* gene in *E. coli* presents numerous complications. 2D protein gel electrophoresis and tandem mass spectroscopy were used to examine the relatively short catalytic lifetime and low activity of PhlD. In an attempt to identify better phloroglucinol synthases, the genetic diversity of *phlD* was explored. Bioinformatics analysis of potential *phlD* candidates in various organisms with significant amino acid sequence identity to our currently used Pf-5 *phlD* were examined. In parallel with these efforts, microbial synthesis of phloroglucinol in *P. fluorescens* was examined. Defined minimal salt medium was formulated to enable high density cultivation of *P. fluorescens* Pf1.162/pJA2.232 in the fermentor.

#### c. Concise Accomplishments

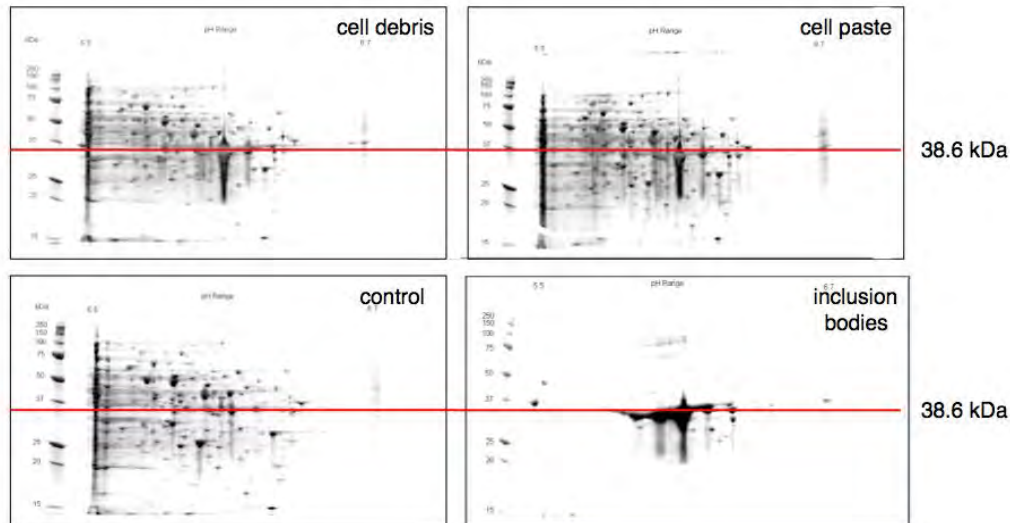
Under optimized resin-based extractive fermentation, *E. coli* PG1/pKIT10.080 synthesized 25 g/L of phloroglucinol. Coupling 2D protein gel electrophoresis and tandem mass spectroscopy, it was determined that heterologously expressing PhlD-encoded phloroglucinol synthase under fermentor-controlled conditions led to significant formation of inclusion bodies. In attempts to identify phloroglucinol synthase alternatives using bioinformatics approach, five active phloroglucinol synthases were identified. High-cell density cultivation of *P. fluorescens* was achieved in fermentation vessels using minimal salts medium. Phloroglucinol production was

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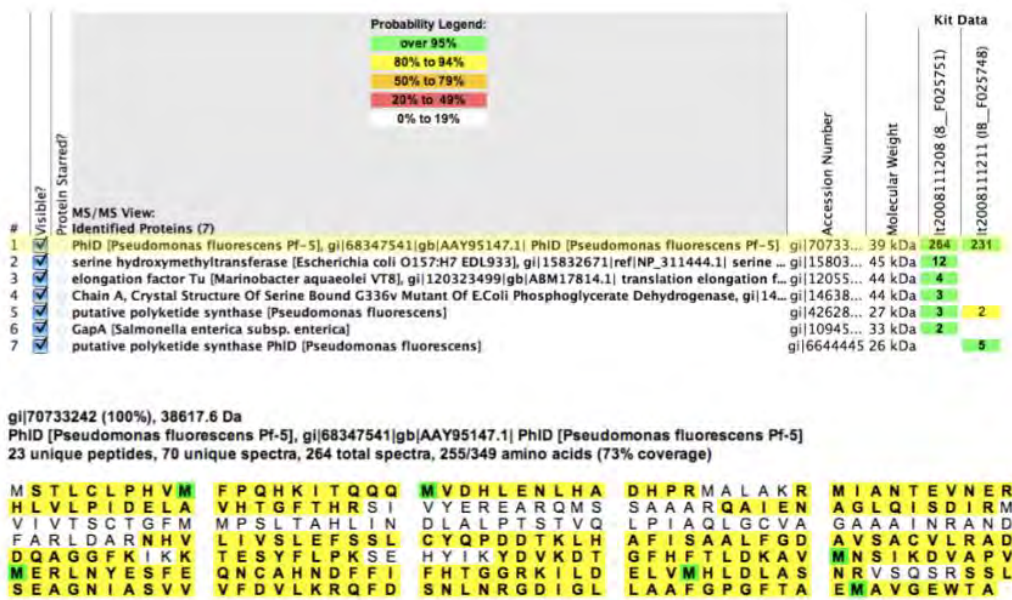
observed by culturing *P. fluorescens* Pf1.162/pJA2.232 under defined fermentor-controlled conditions.

#### d. Expanded Accomplishments

The relative amount of soluble PhID protein produced *in vivo* during the microbial synthesis of phloroglucinol using *E. coli* PG1/pBC2.274 is of particular interest. To this end, a series of 2D protein gel electrophoresis experiments were carried out. Three protein samples were prepared using a 1 L fermentation run of PG1/pBC2.274. The first cell sample (Figure 1, control) was collected at  $OD_{600} = 50$  before induction of phloroglucinol synthase synthesis using IPTG. The fermentation was terminated 6 h post-induction and the cells were harvested. A portion of the cells was washed with HEPES buffer and centrifuged to pellet (Figure 1, cell paste). A third sample was prepared by collecting cell debris after several French Press passages (Figure 1, cell debris). The remaining cells were used to prepare insoluble protein fraction (Figure 1, inclusion bodies). These samples were loaded on 2D protein gels using a pH gradient ranged from 5.5 to 6.7 and an intense dark spot was observed at the intersection of pH 6 and 38 kDa on all the gels. This protein spot was excised from the gel and analyzed by tandem mass spectroscopy with electrospray ionization method. Using the protein identification software Scaffold, the isolated protein fraction was unambiguously identified as phloroglucinol synthase. Therefore, it was concluded that culturing phloroglucinol producing microbe PG1/pBC2.274 led to the formation of a significant amount of inclusion bodies under fermentor-controlled conditions. Attenuate PhID expression level in *E. coli* might lead to a better phloroglucinol synthesizing biocatalyst by alleviate metabolic burden due to the expression of inactive protein.

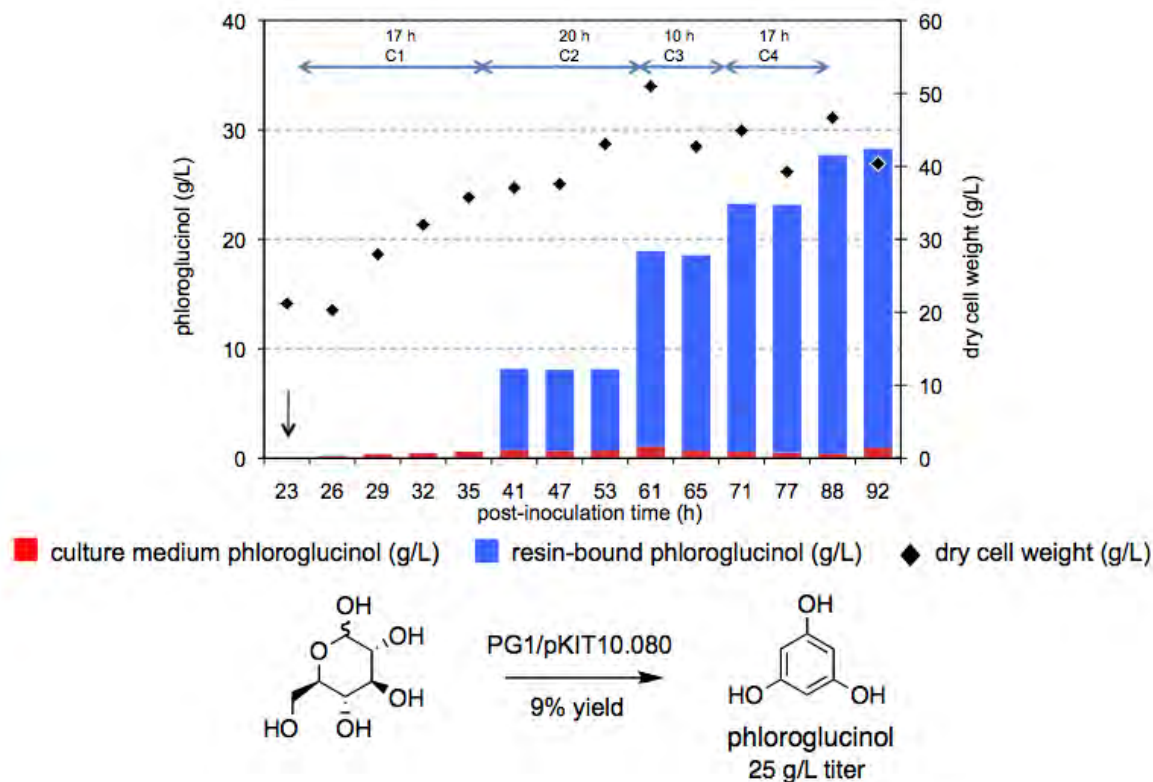


**Figure 1.** 2D protein gel electrophoresis of PG1/pBC2.274 samples.



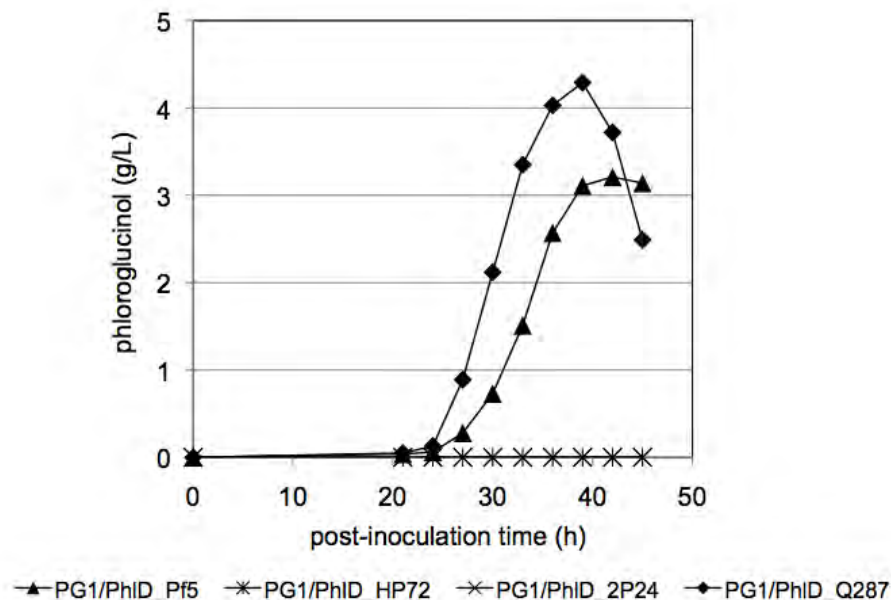
**Figure 2.** PhlD identification using Scaffold.

A series of genetically engineered *E. coli* was developed accordingly. Among these candidates, *E. coli* PG1/pKIT10.080 demonstrated a significant improvement in strain stability and phloroglucinol production under fed-batch fermentor-controlled cultivation. Previously, *in situ*, resin based extractive fermentation was employed to remove toxic substances generated during bacterial culturing. This technology was used once again to remove toxic phloroglucinol from the production tank. Under optimized extractive fermentation conditions, *E. coli* PG1/pKIT10.080 synthesized 25 g/L phloroglucinol in 92 h at a yield of 9% (mol/mol) based on glucose consumed.



**Figure 3.** Resin-based extractive fermentation of *E. coli* PG1/pKIT10.080.

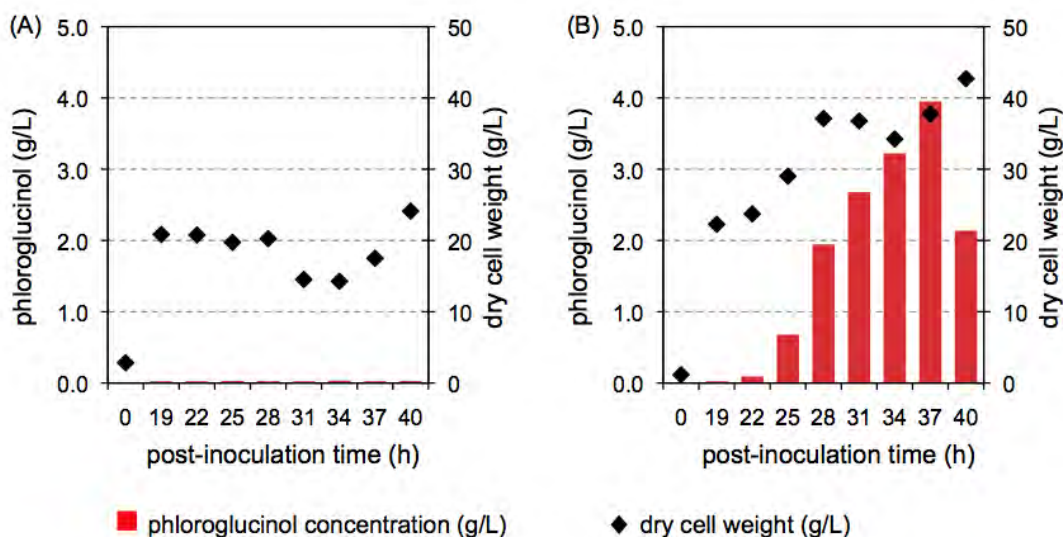
Four full length PhlD sequences have been published to date. PhlD from *Pseudomonas fluorescens* Q2-87 (designated as PhlD<sup>Q2-87</sup>) is 83% identical at the protein sequence level with PhlD<sup>Pf-5</sup>. PhlD from *Pseudomonas fluorescens* HP72 (designated as PhlD<sup>HP72</sup>) is 85% identical at the protein sequence level with PhlD<sup>Pf-5</sup>. PhlD from *Pseudomonas fluorescens* 2P24 (designated as PhlD<sup>2P24</sup>) is 84% identical at the protein sequence level with PhlD<sup>Pf-5</sup>. In addition, approximately 240 partial PhlD sequences are listed in the NCBI GenBank. As with the aforementioned PhlD full length sequences, the partial PhlD sequences are very similar and share 85% or greater sequence identity with PhlD<sup>Pf-5</sup>. Four phloroglucinol synthesizing *E. coli* PG1/PhlD\_Pf5, PG1/PhlD\_HP72, PG1/PhlD\_2P24 and PG1/PhlD\_Q287 were constructed using *de novo* synthesized phloroglucinol synthase candidate genes that were codon-optimized for expression in *E. coli*. *E. coli* PG1/PhlD\_Q287 successfully synthesized 4.3 g/L of phloroglucinol under fed-batch fermentation conditions (Figure 4).



**Figure 4.** Fed-batch fermentation experiments of PhlD candidates.

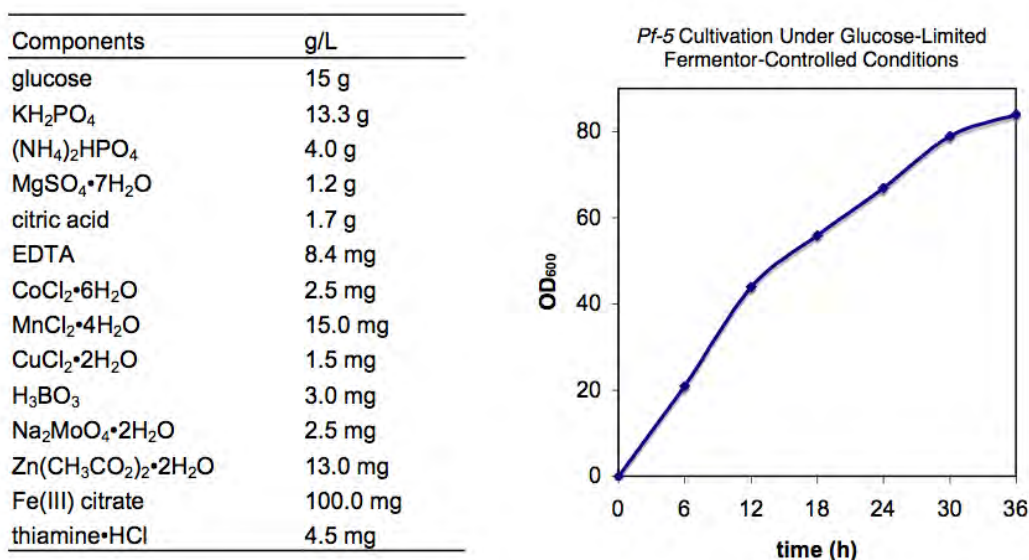
All previous phloroglucinol synthases shared approximately 85% identity at the protein sequence level. During the course of our studies, novel phloroglucinol synthases PhlD<sup>A</sup> and PhlD<sup>B</sup> were identified that share no more than 46% identity at the protein sequence level with all previously isolated phloroglucinol synthases such as the extensively studied *Pseudomonas fluorescens* Pf-5 phloroglucinol synthase (designated as PhlD<sup>Pf-5</sup>). Initial identification of these phloroglucinol synthases followed from bioinformatics analyses. The PhlD<sup>A</sup> protein sequence is only 46% identical with PhlD<sup>Pf-5</sup>. The PhlD<sup>B</sup> protein sequence is only 43% identical with PhlD<sup>Pf-5</sup>. PhlD<sup>A</sup> and PhlD<sup>B</sup> are also very different from one another sharing only 46% identity at the protein sequence level. Unlike the *phlACBDE* gene cluster in which *phlD*<sup>Pf-5</sup> resides in *Pseudomonas fluorescens* Pf-5, *phlD*<sup>A</sup> and *phlD*<sup>B</sup> were not part of a *phlACBDE* biosynthetic gene cluster or any other apparent gene cluster. Genes encoding PhlD<sup>A</sup> and PhlD<sup>B</sup> were synthesized and appraised *in vitro* for enzyme activity in the presence of substrate malonylCoA. Crude cell lysate from PhlD<sup>A</sup> gave specific activity of 0.001 U/mg, while PhlD<sup>B</sup> gave 0.01 U/mg. Plasmid-localized *phlD*<sup>A</sup> and *phlD*<sup>B</sup> were also transformed into an *E. coli* host PG1 and the intact constructs evaluated for synthesis of phloroglucinol under fermentor-controlled conditions (Figure 5). Peak phloroglucinol synthesis of 0.2 g/L was observed for *E. coli* PG1/PhlD<sup>A</sup>. This compares with peak phloroglucinol synthesis of 4.0 g/L for *E. coli* PG1/PhlD<sup>B</sup>.



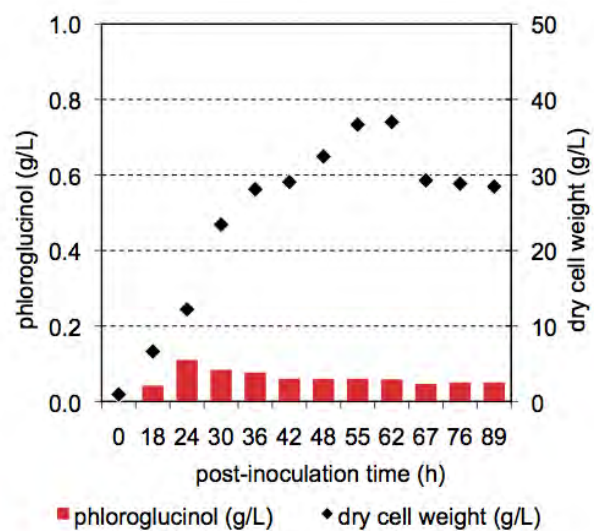


**Figure 5.** Fed-batch fermentation experiments of *E. coli* PG1/PhlD<sup>A</sup> and PG1/PhlD<sup>B</sup>.

In parallel with all these efforts, microbial synthesis of phloroglucinol in *P. fluorescens* was also examined. High cell density cultivation of *P. fluorescens* was achieved in the fermentation vessel using minimal salt medium. The medium formulation is shown in Figure 6. Fed-batch fermentation of wild-type *P. fluorescens* was carried out using this medium formulation and an optical density at 600 nm reached 82 shortly after 36 h cultivation. Cultivation of *P. fluorescens* phloroglucinol synthesizing construct Pf1.162/pJA2.232 under a similar culturing condition produced 0.1 g/L phloroglucinol (Figure 7). This experiment demonstrated the first example of microbial synthesis of phloroglucinol in *P. fluorescens* species under fed-batch conditions using minimal growth medium.



**Figure 6.** Minimal medium formulation for *P. fluorescens* cultivation.



**Figure 7.** Fed-batch fermentation experiment of *P. fluorescens* Pf1.162/pJA2.232.

e. Work Plan

This project is completed.

f. Major Problems/Issues

None.

g. Technology Transfer

None.

h. Foreign Collaborations and Supported Foreign Nationals

None.